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Hala Abdel Salam, Esam S. Al-Malki and Kamal S. Albenasy

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Evaluation of molluscicidal activity and toxicity of some common medicinal plant and chemical synthetic agents against *Biomphalaria alexandrina*, the snail vector of *Schistosoma mansoni* infection

Hala A. Abdel Salam
Department of Zoology, Faculty of Science, Cairo University, Egypt
Email- hala.ali2010@yahoo.com.

Esam S. Al-malki
College of Science, Majmaah University, Majmaah , Saudi Arabia

Email- e.almalki@mu.edu.sa

Kamal S. Albenasy
College of Applied Medical Sciences, Majmaah University, Majmaah, Saudi Arabia
Email-k.albenasy@mu.edu.sa

Abstract- This study aims to evaluate and compare the toxicity and molluscicidal efficacy of three monoterpene oils (linalool, thymol, and eugenol) and artemether, plant derivative agents and atrazine, roundup and miltefosine, chemical synthetic agents against freshwater snails *Biomphalaria alexandrina*; intermediate host of *Schistosoma mansoni*. Following WHO guidelines, the sublethal (LC₀, LC₁₀ and LC₂₅) and lethal (LC₅₀ and LC₉₀) concentrations of tested materials were estimated after 24 hours exposure followed by another of recovery. The recorded results illustrated that all the tested agents had molluscicidal properties and a toxic effect against the examined snails. The most potent molluscicidal agent was atrazine expressed by LC₅₀ of 1.31 ppm and LC₉₀ of 4.07 ppm followed by roundup with LC₅₀ 3.21 ppm and LC₉₀ 12.61 ppm, then miltefosine with LC₅₀ 14.66 ppm and LC₉₀ 24.06 ppm, then artemether with LC₅₀ 18.30 ppm and LC₉₀ 28.40 . Furthermore, the recorded results showed that the molluscicidal activity of the tested agents was arranged according to their lethal concentrations against *B. alexandrina* snails as the following: atrazine > roundup > miltefosine > artemether > thymol > eugenol > linalool. This study highlights plant derivative agents as safe natural molluscicides instead of harmful toxic chemical synthetic agents on non-target organisms and even man. It is recommended that more research studies should be carried out to assess the impact of linalool, thymol, eugenol and artemether in terms of biological, biochemical and molecular parameters of snails as well as their efficacy as schistosomicidal agents against *S. mansoni* worms.

Keywords – Plant molluscicides, Chemical molluscicides, *Schistosoma mansoni* control

I. INTRODUCTION

Fresh water *Biomphalaria alexandrina* snails have a great medical importance as vectors for *Schistosoma mansoni*. *S. mansoni* is a significant etiological agent of intestinal human schistosomiasis [1]. Snail's control is represented one of the most effective method for schistosomiasis control and the best available mean for elimination a link in *Schistosoma* transmission cycle [2]. Generally, the use of molluscicides in snails' control has received particular attention due to the reduction of incidence and prevalence of schistosomiasis [3]. Economic and environmental concerns are increasingly linked to the use of molluscicides that are selectively effective, environmentally friendly, cheap and readily accessible in the affected areas (Bakry et al.,2017) [3].

The use of chemical synthetic molluscicides is hampered by their expensive cost, their toxicity for human and other non-target living organisms. Additionally, they may evoke harmful biological effects on the environment beyond those for which they were originally manufactured [2,3]. In view of these disadvantages, particular renewed interest has been directed to plant molluscicides that are inexpensive, safe, biodegradable, environmentally acceptable agents, readily available and simply applicable in the affected areas especially in the developing countries [4,5]. Molluscicidal activity has been evaluated for more than 1000 plant species [3]. Monoterpenes are a group of biogenic compounds that found in many different higher order plants and give them their unique odorous properties [6]. Monoterpenes have structures which derived from the coupling of two isoprenoid units that are made from isopentylpyrophosphate. These compounds are usually aromatic oils which are frequently found in perfumes and other cosmetics and are often used as food additives and therapeutic medications (Abdelgaleil., 2010) as well as they used as a natural pesticidal and molluscicidal agents [6, 7,8]. Furthermore, artemether is a methyl ether derivative of artemisinin that was extracted from the leaves of the Chinese wormwood plant (*Artemisia annua*) [4]. It possesses a broad spectrum of activities against all species of malaria [9]. Additionally, it has been shown that artemether was confirmed to have a molluscicidal efficacy against intermediate host of *S. mansoni* snails as the result of its bad effect on physiological and biological activity of treated snails [2, 4]. Also, artemether exhibited anti-schistosomal properties against the larval stages and adult worms of human schistosomal species [7, 10].

Based on the mentioned background, the present work aims to assess the molluscicidal efficacy of three monoterpene oils (linalool, thymol and eugenol) and artemether, natural plant molluscicide products in comparison with the molluscicidal activity of two common pesticides; roundup (glyphosate) and atrazine and miltefosine (hexadecylphosphocholine), chemical synthetic molluscicides against *B. alexandrina* snails. Atrazine and roundup are used as herbicides in agricultural areas and in non-agricultural situations to combat weeds and grasses [11]. Whereas miltefosine is a membrane-active drug that was developed as oral medication to treat various parasite species such as different forms of leishmaniasis, *Trypanosoma*, *E. histolytica* and *Schistosoma*. Moreover, it used as anticancer agent [11, 12, 13].

II. MATERIALS AND METHODS

2.1 Experimental snails–

Uninfected adult *B. alexandrina* snails (8-10 mm in diameter) were brought from Schistosomiasis Biological Supply Centre (SBSC) at Medical Malacology Laboratory, Theodor Bilharz Research (Giza, Egypt). Snails were reared in dechlorinated water and fed daily with fresh lettuce leaves (0.015 g/day/snail) plus dried flakes (TetraMin, Hanover, Germany). Snails were maintained under standard laboratory conditions [14].

2.2. Experimental materials–

- 1- The monoterpene oils (linalool, thymol and eugenol) were purchased from Sigma Chemical Company, USA.
- 2-Artemether is an antimalarial agent was obtained in tablet form (Kunming Pharmaceutical Cooperation, PR China) with a documented purity of 99.6%.
- 3- Miltefosine (100 mg) was provided by (Sigma-Aldrich Chemie) and (GmbH, CA 58066-85-6, MW 407.57, Germany).
- 4- Roundup (glyphosate concentration 120 g/l in the form of glyphosateisopropylamine salt 162 g/l). Roundup pesticide was used in the liquid commercial form was supplied by Monsanto Company (St. Louis, MO, USA).
- 5- Atrazine (2-chloro-4-ethylamino-6-isopropylamino-s-triazine, 97.8% purity) was purchased from Cluzeau Info Labo (Ste Foy La Grande, France).

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2.3. Estimation of molluscicidal activity of experimental materials:

The efficacy of the tested materials as molluscicidal agents against adult *B. alexandrina* snails was determined according to WHO (1965) [15]. A stock solution of 1000 ppm from each material (artemether, miltefosine, roundup and atrazine) was prepared according to its active ingredient (concentration/volume). For linalool, thymol and eugenol oils, stock solution (1:1) of each oil was prepared by measuring out 1 ml of each oil and emulsified with Tween-80 of about 0.003 ml or 3 drops from a needle tip [16]. The emulsified oil is then added up to 1 liter of dechlorinated water to form stock solution (1000 ppm). A series of concentrations expressed in terms of part per million (ppm) were prepared from each stock solution that would permit the computation of experimental concentrations LC₅₀ and LC₉₀ [17]. The LC₀ was estimated as 1/10 of the LC₅₀ value [14] (WHO, 1965). Mortality rates were recorded and SPSS was used to computer program under windows. This experiment was carried out by preparing three replicates of gradual concentrations from each stock solution. 30 snails / each replicate were used. The snails were exposed to the tested concentrations for 24 hours, then removed from the experimental concentration, washed with dechlorinated water and they kept in 1 liter of dechlorinated tap water for next 24 hours for recovery (25±2°C).

2.4. Statistical analysis

Analysis of the obtained data statistically analyzed by using the Statistical Package for Social Science (SPSS version 15 package software). Each experiment was replicated three time and the recorded values were expressed as means ± S.E. The examined parameters of the treated and control snails were compared by student “t” test [18]. Statistically significant differences between groups were considered to have a p value of < 0.05.

III. EXPERIMENT AND RESULT

The molluscicidal property of natural tested materials against adult *B. alexandrina* snails after 24 h of exposure followed by another of recovery is presented in Table 1. The recorded data illustrated that artemether had the most toxic effect comparable with studied monoterpene oils, i.e. the least lethal concentration, LC₅₀ (18.30ppm) and LC₉₀ (28.40ppm) values were recorded for artemether with slope function value of 1.24. While, the highest LC₅₀ and LC₉₀ were 36.00ppm and 56 ppm, respectively with slope function value of 1.49 was recorded for linalool. Furthermore, it was noticed that the molluscicidal efficacy of natural studied materials had the following decreasing order according to their lethal and sublethal concentrations: artemether > thymol > eugenol > linalool.

Table -1 Experiment Result

Experimental material	LC ₀ ppm	LC ₁₀ ppm	LC ₂₅ Ppm	LC ₅₀ ppm	LC ₉₀ ppm	Slope function
Linalool	4.00 ^a	22.00 ^a	28.00 ^a	36.00 ^a	56.00 ^a	1.49 ^a
Thymol	2.10 ^b	9.45 ^b	15.94 ^b	21.90 ^b	34.06 ^b	1.65 ^b
Eugenol	2.55 ^c	12.05 ^c	19.88 ^c	26.77 ^c	48.02 ^c	2.05 ^c
Artemether	1.83 ^b	5.64 ^d	10.50 ^d	18.30 ^d	28.40 ^d	1.24 ^d

Values means with the same letter for each parameters are not significantly different, otherwise they do (P<0.05).

Table 1 show the sublethal and lethal concentrations of natural experimental agents to illustrate their molluscicidal activity, where artemether has the lowest sublethal and lethal concentrations comparable with monoterpene agents.

These recoded results are in agreement with the results of Mossalem et al. (2013) [4] that cleared the efficacy of artemether as a toxic molluscicidal agent against *B. alexandrina* snails with LC₅₀ of 21.06 ppm and LC₉₀ of 32.69 ppm after 24 hours exposure. Similarly, after the same time period of exposure, Osman et al. (2019) [2] verified that LC₅₀ and LC₉₀ of artemether on *B. alexandrina* snails were 16.88 ppm and 27.97 ppm, respectively.They attributed

the molluscicidal toxicity of artemether against snails due to immunological alterations that represented by negative effect on hemocytes, i.e. artemether evoked their degenerative changes and fragmentation. This distribution in snails' tissues induced by artemether that may deteriorate the biological and physiological activities of exposed snails. Several authors also reported that monoterpenes oils included eugenol, linalool and thymol had acute toxicity on schistosomiasis and fascioliasis vector snails [6, 19, 20]. Moreover, the results of El-Din (2006) [19] showed that thymol was the potent monoterpene oil against *B. alexandrina* snails at least LC₅₀ (22 ppm) and LC₉₀ (34 ppm) followed by eugenol (LC₅₀ of 28 ppm and LC₉₀ of 48 ppm) then linalool (LC₅₀ of 34 ppm and LC₉₀ of 56 ppm). Similarly, Hamdi et al. (2008) [6] showed that according to lethal concentrations (LC₅₀ and LC₉₀), thymol was the most effective monoterpene followed by eugenol, then linalool against *Bulinus truncatus* (intermediate hosts of *S. haematobium*) and other aquatic snail species (*Planorbis planorbis*, *Helisoma dur*, *Physa acuta*, *Cleoparta bulimoides* and *Melanoides tuberculata*) that naturally associated with *B. truncatus*. Thymol exhibited high molluscicidal properties against *Schistosoma*, *Fasciola* snails and other harmful terrestrial snails [21,22] Monoterpene oils have molluscicidal activity as the result of their negative effect on the snails' physiology and biochemistry activities [6, 19, 23]. Additionally, it was proved that monoterpene oils (thymol, eugenol and linalool) evoked an inhibitory effect in the level of some enzymes such as transaminases activity in the haemolymph and tissues of exposed *Schistosoma* vector snails [6]. Also, the studies showed that monoterpene oils had other useful activities such as eugenol was larvicidal; antimicrobial antioxidant and anti-inflammatory against harmful snails [23]. Thymol had anti-inflammatory, fungicidal and bactericidal activities [24] , furthermore, it was often used as a rapidly decaying, toxic pesticide [25].

Regarding the chemical synthetic agents, the data obtained in Table 2 clearly indicated that atrazine was the most potent synthetic material with the lowest sublethal and lethal concentrations against *B. alexandrina* snails followed by roundup then miltefosine. Statically significant differences were recorded (P < 0.001). In comparison with the molluscicidal effect of the studied plant origin agents, the results detected that chemical synthetic agents were more toxic on *B. alexandrina* snails than the natural agents. The present study demonstrated that all the studied tested materials had a molluscicidal effect against intermediate host of *S. mansoni*, based on the standardized WHO method (1965) [15] , the median lethal concentration (LC₅₀) must not exceed 100 ppm for any molluscicidal material. The lowest LC₅₀ was 1.31ppm for atrazine, whereas the highest value was 36.00 ppm for monoterpene oil linalool. According to the recorded values of sublethal and lethal concentrations in the present work, this order was noticed: atrazine > roundup > miltefosine > artemether > thymol > eugenol > linalool.

Table -2 Experiment Result

Experimental material	LC ₀ ppm	LC ₁₀ ppm	LC ₂₅ Ppm	LC ₅₀ ppm	LC ₉₀ ppm	Slope function
Miltefosine	1.48 ^a	3.55 ^a	8.21 ^a	14.66 ^a	24.05 ^a	1.45 ^a
Roundup	0.64 ^b	0.88 ^b	2.03 ^b	3.21 ^b	12.61 ^b	2.40 ^b
Atrazine	0.21 ^c	0.35 ^c	3.19 ^c	1.31 ^c	4.07 ^c	2.21 ^c

Values means with the same letter for each parameters are not significantly different, otherwise they do (P< 0.05).

Table 2 show the sublethal and lethal concentrations of chemical synthetic experimental agents to illustrate their molluscicidal activity, where atrazine was most potent molluscicidal agent.

The recorded results agree with Barky *et al.* (2012) [11] who detected that atrazine was more efficient molluscicidal agent to freshwater snails than roundup. They reported a considerable killing effect of pesticides; roundup and atrazine to *B. alexandrina* snails with LC₅₀ of 3.15 and 1.25 ppm, and LC 90 of 12.60 and 4.75 ppm, respectively. The authors determined also the sublethal dose LC₁₀ (0.33 ppm for roundup and 0.84 ppm for atrazine). Whereas, the study of Omran and Salama (2013) [26] showed that the sublethal doses LC₁₀ were 4.02 and 10.1 ppm for roundup and atrazine, respectively. Their recorded lethal doses LC₅₀ values were 41.6 and 101.16 ppm, respectively on *B. alexandrina* after 24 hours of exposure followed by another of recovery. The toxicity of these herbicides against *Biomphalaria* snails is genotoxic leading to DNA damage which resulted from disturbance of antioxidant defensive system by increasing of free radical life span in exposed snails [27]. In addition to genotoxicity effect of roundup and atrazine, Bakry *et al.* (2012) [11] attributed their molluscicidal activity against *B. alexandrina* as the result of their adverse effects on metabolism and reproduction of snails. Furthermore, the results of Omran and Salama (2013) [26] proved that both of roundup and atrazine are considered endocrine disrupters due to their efficacy to alter the endocrine functions and cause cellular toxicity and histological alterations of *B. alexandrina* snails including deformation of sperms and oocytes. Regarding with molluscicidal effect of miltefosine against *B. alexandrina*, Eissa *et al* (2011) [11] proved that miltefosine is a potential promising molluscicidal

compound against *S. mansoni* vector, *B. alexandrina* (LC₅₀ of 7.5 ppm and LC₉₀ of 9.9 ppm) and against *S. haematobium* vector, *B. truncates* snails (LC₅₀ of 2.6 ppm and LC₉₀ of 4.2 ppm). Meanwhile, the lethal concentrations of miltefosine against *B. alexandrina* were 13.80 ppm and 24.40 ppm for LC₅₀ and LC₉₀ as recorded by Osman *et al.* (2019) [2]. Miltefosine induced considerable morphological alterations of *B. alexandrina* such as extensive damage of the tentacles and the cilia at the foot as observed by scanning electron microscopy of exposed snails (Eissa *et al.*, 2011a). Additionally, its toxicity as molluscicidal agent resulted from its negative effect on immunological aspects including fragmentation and degenerative of haemocytes [2].

Despite the efficacy of atrazine, roundup and miltefosine as the most potent molluscicidal agents comparable with plant natural molluscicidal agents linalool, thymol, eugenol and artemether against the examined snails, these synthetic materials may occasionally cause sublethal effects in various living organisms. So, the application of these synthetic products should be limited on their use as authorized molluscicides in the control program of snail vectors of schistosomiasis due to their biocidal effects on plants and animals, besides, their ecological toxicity, genotoxicity and carcinogenic impact on non-target organisms and even man, in addition to their expansive cost especially in developing countries [20]. Atrazine and roundup have the ability to accumulate in tissues of fish, phytoplankton and other aquatic invertebrates due to their physical, chemical properties and high water solubility as well as their extensive usage as grasses and weeds pesticides in agricultural applications and in non- agricultural situations [11]. Their bioaccumulation caused in turn harmful toxic effects for aquatic ecosystems through influencing on the reproduction of aquatic flora and fauna and interfering in the food chains of many species which in turn impacts on the community structure as a whole as documented by several researches [11, 28,29]. Roundup as a synthetic pesticide had an inhibitory effect on reproduction and development of arthropods, molluscan snails, fish, amphibians and plants by evoking biochemical, physiological and immunological alterations (Kielak *et al.*, 2011). Similarly, the studies detected that atrazine induced serious environmental and human health risks and caused toxic effects to aquatic species [29]. It is an endocrine deactivate agent that deteriorates reproductive capacity of humans and animals [30]. Atrazine induced a decrease in oocyte meiotic maturation *in vitro* [31]. Furthermore, it altered the growth, behavior, reproduction and survival of algal, aquatic plants, invertebrates, zooplankton and fish. In addition to, it reduced cellular metabolism and deteriorate the antioxidant activity of aquatic organisms [29, 32]. As regards to toxic effect of miltefosine on non-target organisms, the results of Eissa *et al.* [12] demonstrated that the molluscicidal activity of miltefosine as a synthetic molluscicide agent had a broad biocide activity that might make it inappropriate for snail control. Additionally, the previous studies [33,34] showed that miltefosine is teratogenic agent and its action in treatment as anti-leishmanial drug was associated with severe side effects. Generally, human exposure to chemical synthetic pesticides or other synthetic agents poses a higher health risk than exposure to genetically modified foods [35]. So, the use of natural alternatives is represented a good solution that can reduce the side effects of impact of chemical synthetic molluscicides.

IV.CONCLUSION

The present study provides an information on the efficiency of artemether and monoterpene oils (linalool, thymol, and eugenol) as natural molluscicidal agents against *S. mansoni* snails. These tested agents were recommend as promising and safe natural molluscicide agents, besides they can be effectively implemented using simple techniques acceptable for developing countries. These were also mentioned as possible eco-friend molluscicides and alternatives to the currently available expensive and environmentally destructive molluscicides (atrazine, roundup and miltefosine). The obtained results could be useful to the National *S. mansoni* control programs and the public health service, therefore further research studies are warranted to evaluate the efficacy of artemether and linalool, thymol and eugenol on biological, biochemical and molecular parameters of snails as well as their effects on *S. mansoni* worms as schistosomicidal agents. Also, from the results of the present study, it showed that *B. alexandrina* snails may be used as a bioindicator of water pollution and pesticides toxicity in the surrounding water.

REFERENCES

- [1] M.M .Abdel- Ghaffar, E. S.Abdel- Gawad,. I M Moharm, O F Sharaf, M T Badr and AF Ibrahim, "Parasitological and histopathological effects of some antischistosome drugs in *Schistosoma mansoni*-infected mice". *Menoufia M. J.*, vol.30, pp 1193–1202, 2017.
- [2] G Y. Osman,. H H. Abdel-Azeem, M F. El Garhy and K S. Al Benasy " Sublethal toxicity of miltefosine and artemether to molecular aspect of *Biomphalaria alexandrina* snails", *Chem. Bio. Phy. Sci. Sec. B.* Vol.9, No.3, pp 405-413, 2019.
- [3] FA Bakry, M F. El-Garhy, M. Abd El-Atti and M T. Atwa "Effects of the extracts of *Euphorbia pulcherima* and *Atriplex nummularia* on the infectivity of *Schistosoma haematobium* to *Bulinus truncatus* snails", *ABM J.*, Vol 1, No.2, pp. 30-41, 2017.

- [4] H. S. Mossalem, H. Abdel-Hamid, and N. A. El-Shinnawy, "Impact of artemether on some histological and histochemical parameters in *Biomphalaria alexandrina*", *African J. Pharm. Pharmacol.*, Vol. 267, No.31, pp. 2220-2230,2013.
- [5] GY. Osman, A H. Mohamed., S K .Sheir., S E. Hassab EL-Nabi and S. A. Allam, (2014) "Molluscidal activity of mirazid on *Biomphalaria alexandrina* snails: biological and molecular studies". *Int. J. Advan. Res.*, Vol. 2 , No. 2, pp. 977-989,2014.
- [6] S A Hamdi, A.M.A Sakran and H A Adel Salam "Molluscicidal activity of monoterpenes on *Bulinus truncates* and some naturally associated snails". *J. Egypt . Ger. Soc. Zool.*, Vol .56D, pp. 211-224,2008.
- [7] S.A.M. Abdelgaleil "Molluscicidal and insecticidal potential of monoterpenes on the white garden snail, *Theba pisana* (Muller) and the cotton leafworm, *Spodoptera littoralis* (Boisduval)", *Appl. Entomol. Zool.*, Vol.45 No. 3,pp. 425–433, 2010.
- [8] L. P L A. Pereira, C G R Edilene, C. A Maria, P. B.S Daniella., O. S.A., Fernanda Felipe B.A, A.José, L Costa., et al. "Essential oils as molluscicidal agents against schistosomiasis transmitting snails" - a review. *Acta Trop.*, Vol 209, 10548, 2020.
- [9] Y.Y. Tu, "The discovery of artemisinin (qinghaosu) and gifts from Chinese medicine". *Nat. Med.*, Vol.17, No.10, pp. 1217-1220,2011.
- [10] Yi-Xin, L.,W Wei, L. Yue-Jin, J. Zu-Liang, W. Hui, W Wei, and H. Yi-Xin, (2014): New Uses for Old Drugs: The tale of Artemisinin derivatives in the elimination of Schistosomiasis *Japonica* in China. *Molecules*, Vol. 19, pp. 15058-15074, 2014.
- [11] F.A.Barky F.A, HA. Abdel Salam, M.B. Mahmoud, S.A.H Hamdi, "Influence of Atrazine and Roundup pesticides on biochemical and molecular aspects of *Biomphalaria alexandrina* snails. Pest". *Biochem. Physiol.*, Vol. 104,pp. 9–18, 2012.
- [12] MM Eissa. S. El Bardicy and M. Tadros, "Bioactivity of miltefosine against aquatic stages of *Schistosoma mansoni*, *Schistosoma haematobium* and their snail hosts, supported by scanning electron microscopy", *Paras. Vect.*, Vol.4,pp. 11-18.
- [13] N. El Deeb and F. A. Bakry "Antischistosomal and immune-biochemical effect of miltefosine and plant extraction infected albino mice with *Schistosoma mansoni* ". *IJBPAS*, Vol. 6 No. 4, pp. 1-11.
- [14] M.A. El-Emam, M. A. and F.A. Ebeid, "Effect of *Schistosoma mansoni* infection starvation and molluscicides on acid phosphatase transaminase and total protein in tissues and hemolymph of *B. alexandrina*". *J. Egypt. Soci. Parasitol.*, 19: 139-147,1989.
- [15] WHO "Molluscicide screening and evaluation". *Bull. WHO*,pp. 33:5675 -5681,1964.
- [16] O.O. Anyaele and A.S. Amusan A.S. "Toxicity of hexanolic extract of *Demmettia tripetala* (G), Baxer on larvae of *Aedes aegyptica*". *Afr. J. Biomed. Res.*, Vol. 6,pp. 49-53, 2003.
- [17] JT Litchfield and F. Wilcoxon F. "A simplified method of evaluating dose effect experiments. *J. Pharmacol. Exp. Therap.*,pp.96:99,1949..
- [18] A. Goldstein A. "Biostatistics: An introductory text1964.. Macmillan, New York, pp. 51,1964.
- [19] AT El-Din , "Molluscicidal effect of three monoterpenes oils on schistosomiasis and fascioliasis vector snails in Egypt". *J Egypt Soc Parasitol.*, Vol. 36 No. 2,pp. 599-612, 2006..
- [20] M.A.Hamed, "Strategic control of schistosome intermediate host". *Asian J. Epidemiol.*,pp. 1-16,2010.
- [21] M.A. Radwan, S.R. El-Wakil,S.A Mohamed, S.A. and S.M. Sherby, "Potential Schistosomiasis snail vector, *Biomphalaria alexandrina*". *Ecotox Environ Saf.* Vol. 71, pp. 889-894,2008.
- [22] O.M. Mustafa "Toxicity of thymol on the ultra-scanning structure of skin and digestive gland proteins of the two slugs *Limax maximus* and *Lehmannia marginata*. *Egypt J Hosp. Med.*, 71 (6): 3405-3415,2018.
- [23] P R Barros Gomes, J.Batista Reis , R. Pêsoa Fernandes ,V E. Mouchrek Filho, A. Gouveia de Souza, M. Alves Fontenele, A. Caetano da Silva "Toxicity and molluscicidal activity of the essential oil *Pimenta dioica* against the snail *Biomphalaria*". *Rev. peru. biol.* Vol. 26 No.1,pp. 102 – 110,2019.
- [24] P.Ferreira, L.G.Geraldo, D. Sthefane, D'ávila, C. Elisabeth and C. de Almeida Bessa1, "The influence of thymol+DMSO on survival, growth and reproduction of *Bradybaena similaris* (Mollusca: Bradybaenidae) ". *ZOOL.* Vol.28 No. 2,pp 145–150, 2011.
- [25] G. Nieto, "Biological activities of three essential oils of the Lamiaceae family" *Medicines*, Vol. 4 No.3, .pp.63-70, 2017.
- [26] N.E. Omran and W.M. Salama W.M., "The endocrine disruptor effect of the herbicides atrazine and glyphosate on *Biomphalaria alexandrina* snails". *Toxicol. Ind. Heal.*, pp.1-10, 2013.
- [27] MA. Mohamed, M.A. El-Emam, G.Y. Osman, H. Abdel-Hamid and E.M.A. Rasha E. M. A. "Biological and biochemical responses of infected *Biomphalaria alexandrina* snails with *Schistosoma mansoni* post exposure to the pesticides Basudin and Selcron and the phytoalkaloid Colchicine". *J Evolutionary Biol. Res.*, Vol. 4 No. 2, pp24-32, 2012.
- [28] K.Kielak .C. Sempruch, H.Mioduszezwska J. Klocek and B. Leszczyn, "Phytotoxicity of roundup ultra 360 SL in aquatic ecosystems: biochemical evaluation with duckweed (*Lemna minor* L.) as a model plant, Pest". *Biochem. Physiol*,Vol.99237,pp243-250.
- [29] F. P. de Albuquerque , J. L. de Oliveira, V. Moschini-Carlos and L. F. Fraceto, "An overview of the potential impacts of atrazine in aquatic environments: Perspectives for tailored solutions based on nanotechnology", *Sci. Total Envir.* Vol. 700 ,pp-1-9,2020.
- [30] X.Chen, J.Wang J,H. Zhu, J. Ding and Y. Peng , "Proteomics analysis of *Xenopus laevis* gonad tissue following chronic exposure to atrazine". *Environ.Toxicolo.Chemi.*Vol. 34No.8, pp 1770-1777, 2015.
- [31] B Yuan B, L Shuang, J. Yong-Xun., Z. Ming-Jun., Z. Jia-Bao Z.and K. Nam- Hyung K., "Toxic effects of atrazine on porcine oocytes and possible mechanisms of action". *PLoS ONE* Vol.,pp12-15.
- [32] A Stara, A Kouba, J. Velisek, J. "Biochemical and histological effects of subchronic exposure to atrazine in crayfish *Cherax destructor*". *Chem. Biol. Interact.*Vol. 291, pp. 95–102, 2018.
- [33] H Sindermann and J Engel J, "Development of miltefosine as an oral treatment for leishmaniasis", *Trans. R. Soc. Trop Med. Hyg.*, Vol. 100 No.1 pp S17-S20.
- [34] SK Bhattacharya,PK Sinha, S Sundar, CP Thakur, TK Jha, K Pandey, VR Das, N Kumar, C Lal, N Verma and VP Singh et al. "Phase 4 trial of miltefosine for the treatment of Indian visceral leishmaniasis", *J. Infect. Dis.*Vol. 196 No. 4, pp.591-598, 2007.
- [35] B.Talyn, R Lemon, M Badoella, D Melchiorre, M Villalobos., R.Elias, M.Muller, M Santos.and E Melchiorre, "Roundup, but not roundup-ready corn, increases mortality of *Drosophila melanogaster*". *Toxic.* Vol.7, pp. 28-38, 2019.